Meta-analyses of genome-wide association studies identify multiple loci associated with pulmonary function

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Spirometric measures of lung function are heritable traits that reflect respiratory health and predict morbidity and mortality. We meta-analyzed genome-wide association studies for two clinically important lung-function measures: forced expiratory volume in the first second (FEV_1) and its ratio to forced vital capacity (FEV_1/FVC), an indicator of airflow obstruction. This meta-analysis included 20,890 participants of European ancestry from four CHARGE Consortium studies: Atherosclerosis Risk in Communities, Cardiovascular Health Study, Framingham Heart Study and Rotterdam Study. We identified eight loci associated with FEV_1/FVC (*HHIP, GPR126, ADAM19, AGER-PPT2, FAM13A, PTCH1, PID1* and *HTR4*) and one locus associated with FEV_1 (*INTS12-GSTCD-NPNT*) at or near genome-wide significance ($P < 5 \times 10^{-8}$) in the CHARGE Consortium dataset. Our findings may offer insights into pulmonary function and pathogenesis of chronic lung disease.

Pulmonary function is an easily measurable and reliable index of the physiological state of the lungs and airways[1]. Pulmonary function also predicts mortality in the general population, even among people who have never smoked (never-smokers) who have only modestly reduced pulmonary function and no respiratory symptoms[2,3]. The peak level of pulmonary function attained in early adulthood and its subsequent decline with age are likely influenced by genetic and environmental factors. Tobacco smoking is a major environmental cause of accelerated decline in pulmonary function with age. Other inhaled pollutants also appear to contribute. Familial aggregation studies suggest a genetic contribution to lung function, with heritability estimates exceeding 40%[4,5], but little is known about the specific genetic factors involved. A relatively uncommon deficiency of α1-antitrypsin is the only established genetic risk factor for accelerated decline in pulmonary function and for development of chronic obstructive pulmonary disease (COPD), especially in smokers[4,6]. However, α1-antitrypsin accounts for little of the population variability in pulmonary function[4]. Candidate gene studies suggest that other genetic variants may influence the time course of pulmonary function and its decline in relation to smoking, but these putative genetic risk factors remain unknown[4].

Forced expiratory volume in the first second (FEV_1) and its ratio to forced vital capacity (FEV_1/FVC) are two clinically relevant pulmonary function measures. Although both FEV_1 and FVC are influenced by lung size and can be reduced by restrictive lung diseases, obstructive lung disease leads to proportionately greater reduction

in FEV_1 than FVC. Therefore, reduced FEV_1/FVC, an indicator of airflow obstruction that is independent of lung size, is the primary criterion for defining an obstructive ventilatory defect[1]. Whereas low FEV_1/FVC indicates the presence of airflow obstruction, FEV_1 is used to classify the severity and follow the progression of obstructive lung disease over time[5,7,8].

The first genome-wide association study (GWAS) for pulmonary function evaluating 70,987 SNPs in about 1,220 Framingham Heart Study (FHS) participants revealed no genome-wide significant loci[9]. Recently, a GWAS of FEV_1/FVC using 2,540,223 SNPs in 7,691 FHS participants identified several SNPs on chromosome 4q31 near *HHIP* with genome-wide significance[10]. A GWAS of COPD[11] also implicated the *HHIP* region along with *CHRNA3-CHRNA5* on chromosome 15, a region previously associated with nicotine dependence[12,13].

We conducted meta-analyses of GWAS results for a cross-sectional analysis of pulmonary function (FEV_1/FVC and FEV_1) in 20,890 individuals of European ancestry from four Cohorts for Heart and Aging Research in Genomic Epidemiology (CHARGE) Consortium[14] studies: Atherosclerosis Risk in Communities (ARIC), Cardiovascular Health Study (CHS), FHS and Rotterdam Study (RS-I and RS-II). Given that cigarette smoking is a major risk factor for pulmonary function decline, we conducted meta-analyses with adjustment for smoking status and quantity (pack-years), and in subgroups of those who have ever smoked (ever-smokers) and never-smokers. Loci meeting genome-wide significance and other selected high-signal hits were evaluated for replication with the SpiroMeta consortium, an independent consortium

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Table 1 Characteristics of cohort participants in the CHARGE consortium at the time of pulmonary function assessment

Cohort, number	% male	Age (y)		Height (m)		BMI (kg/m^2)		Current smoking			Former smoking			Pulmonary function			
								%	Pack-years		%	Pack-years		FEV$_1$ (ml)		FEV$_1$/FVC (%)	
		Mean	s.d.	Mean	s.d.	Mean	s.d.		Mean	s.d.		Mean	s.d.	Mean	s.d.	Mean	s.d.
ARIC 7,980	47.2	54.3	5.7	1.69	0.09	27.0	4.9	25.0	35.8	20.3	35.2	24.1	21.4	2,941	777	73.6	8.0
CHS 3,140	39.0	72.3	5.4	1.65	0.09	26.3	4.4	10.8	45.1	25.4	40.0	29.9	26.4	2,116	659	70.5	10.5
FHS 7,694	46.1	51.9	14.6	1.69	0.10	27.3	5.3	15.3	32.1	23.8	38.5	19.0	19.3	3,038	944	75.1	8.0
RS-I 1,224	45.4	74.5	5.6	1.67	0.09	27.4	4.0	11.7	40.1	23.4	58.8	24.7	22.5	2,320	728	73.1	8.3
RS-II 852	44.7	67.2	6.3	1.68	0.09	27.7	4.1	14.0	37.4	21.4	52.3	21.5	22.0	2,716	782	75.8	9.1

Genotyping platforms used: ARIC, Affymetrix GeneChip SNP Array 6.0; CHS, Illumina Human 370CNV BeadChip; FHS, Affymetrix GeneChip Human Mapping 500K Array and 50K Human Gene Focused Panel; RS-I, Illumina Infinium II HumanHap 550K Single and Duo Bead Chips; RS-II, Illumina Infinium II HumanHap 500K Dup and 610K Quad Bead Chips. FEV$_1$, forced expiratory volume in one second; FEV$_1$/FVC, ratio of forced expiratory volume in one second to forced vital capacity.

having a combined GWAS sample size of 20,288 participants of European ancestry as described in the companion paper[15].

RESULTS

Meta-analyses of CHARGE genome-wide association results

Meta-analyses for FEV$_1$/FVC and FEV$_1$ were conducted using approximately 2,534,500 SNPs in 20,890 CHARGE participants of European ancestry (n = 7,980 from ARIC, n = 3,140 from CHS, n = 7,694 from FHS, n = 1,224 from RS-I and n = 852 from RS-II) and in subgroups of ever- (n = 11,963) and never-smokers (n = 8,927). Characteristics of the cohort participants are presented in **Table 1**. We applied genomic control, although cohort-specific genomic inflation factors (λ_{gc}) were low (for FEV$_1$/FVC ranging from 1.00 (RS-I and RS-II) to 1.05 (ARIC) and for FEV$_1$ ranging from 1.01 (RS-II) to 1.05 (FHS)), suggesting minimal population stratification. The meta-analysis λ_{gc} was 1.04 for FEV$_1$/FVC and 1.03 for FEV$_1$ in all participants. Quantile-quantile (Q-Q) plots show large deviations between observed and expected P values for high-signal SNPs in analyses of FEV$_1$/FVC and FEV$_1$ in all participants (**Supplementary Fig. 1a,b**), FEV$_1$/FVC in never-smokers (**Supplementary Fig. 2a**), and FEV$_1$ in ever-smokers (**Supplementary Fig. 3c**). Genome-wide significant associations ($P < 5 \times 10^{-8}$) were found for multiple SNPs in each of these analyses (see **Fig. 1a,b** for overall analyses and **Supplementary Fig. 2b,d** and **Supplementary Fig. 3b,d** for analyses stratified by ever or never smoking). The top 2,000 SNPs associated with each measure, FEV$_1$/FVC and FEV$_1$, beyond genome-wide significance ($P > 5 \times 10^{-8}$) are presented in **Supplementary Table 1**.

For FEV$_1$/FVC, genome-wide significant associations were seen for 119 SNPs at seven loci (**Supplementary Table 2**). The SNP with the smallest P value, rs1980057 ($P = 4.90 \times 10^{-11}$), is located on chromosome 4q31.22 81 kb away from the 5′ end of *HHIP*. There were 27 other genome-wide significant SNPs in the *HHIP* region (**Fig. 2a**). Additionally, 69 genome-wide significant SNPs were located in or near the 3′ end of *GPR126* on chromosome 6q24.1, with the top SNP (rs3817928) having $P = 2.60 \times 10^{-10}$ (**Fig. 2b**). Fifty-nine of these 69 *GPR126* SNPs were associated with FEV$_1$/FVC at genome-wide significance among never-smokers (**Supplementary Table 2**). Seven SNPs on chromosome 5q33.3 located in *ADAM19* (**Fig. 2c**), two correlated SNPs on chromosome 6p21.32 ($r^2 = 0.66$, **Fig. 2d**) located in two genes (*AGER* and *PPT2*), four SNPs on chromosome 4q22.1 near the 5′ end of *FAM13A* (**Fig. 2e**), two SNPs on chromosome 9q22.32 in *PTCH1* (**Fig. 2f**), and six SNPs on chromosome 2q36.3 near the 3′ end

of *PID1* (**Fig. 2g**) were also significantly associated with FEV$_1$/FVC in all participants. SNPs in *AGER*, *PPT2*, *PTCH1* and *PID1* had minor allele frequencies (MAFs) between 4% and 10%, whereas all other significantly associated SNPs had MAFs exceeding 10%. Absolute values of β (per-allele change) for FEV$_1$/FVC ranged from 0.44% to 1.14%. The directions of β were consistent across the CHARGE cohorts for all genome-wide significant SNPs except for the *GPR126* SNPs noted in **Supplementary Table 2**. A borderline significant association ($P = 5.37 \times 10^{-8}$, MAF = 0.42, $\beta = -0.43$) with FEV$_1$/FVC was noted for the chromosome 5q33.1 SNP rs11168048 in *HTR4* (**Fig. 2h**).
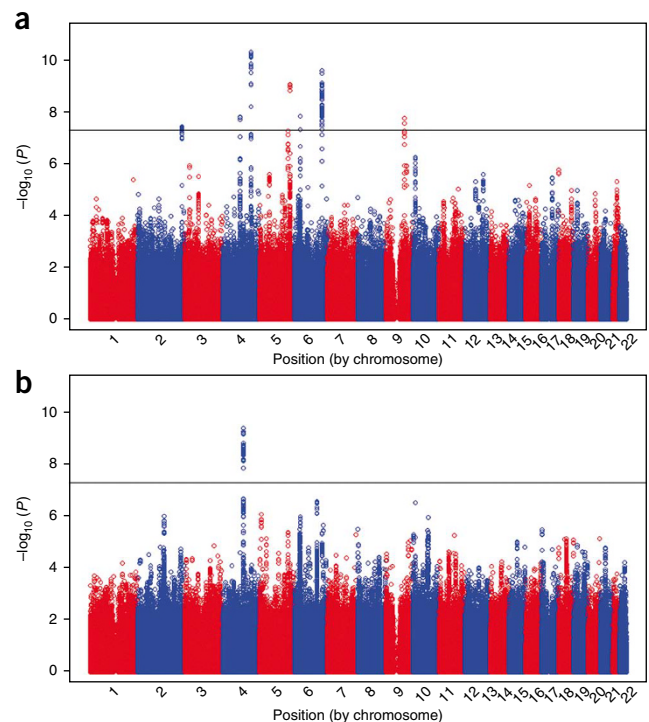
Figure 1 Meta-analyses of approximately 2,534,500 SNPs tested for association with pulmonary function measures in all participants from the CHARGE Consortium. (**a**) FEV$_1$/FVC. (**b**) FEV$_1$. The Manhattan plots (also known as $-\log_{10}$ (P) association plots) show the chromosomal position of SNPs exceeding the genome-wide significance threshold ($P < 5 \times 10^{-8}$ as indicated by the solid black line).
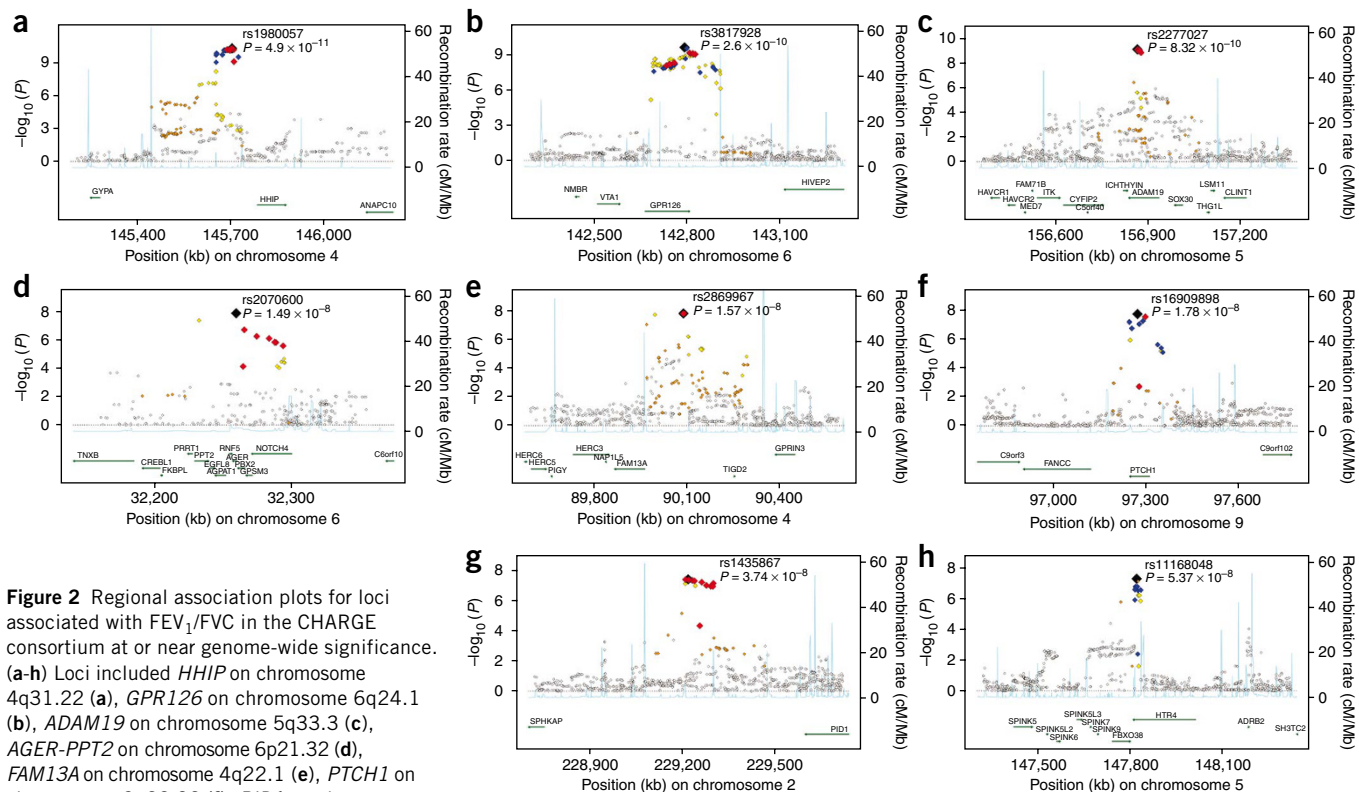
Figure 2 Regional association plots for loci associated with FEV$_1$/FVC in the CHARGE consortium at or near genome-wide significance. (**a**-**h**) Loci included *HHIP* on chromosome 4q31.22 (**a**), *GPR126* on chromosome 6q24.1 (**b**), *ADAM19* on chromosome 5q33.3 (**c**), *AGER-PPT2* on chromosome 6p21.32 (**d**), *FAM13A* on chromosome 4q22.1 (**e**), *PTCH1* on chromosome 9q22.32 (**f**), *PID1* on chromosome 2q36.3 (**g**) and *HTR4* on chromosome 5q33.1 (**h**). For each locus, correlations between the target SNP (the SNP with the lowest *P* value, depicted in black) and other SNPs in the region are depicted in red when $r^2 = 1$, blue when $0.8 \leq r^2 < 1$, yellow when $0.5 \leq r^2 < 0.8$, orange when $0.2 \leq r^2 < 0.5$ and white when $r^2 < 0.2$. The r^2 values were based on the HapMap CEU population. Gene annotations are shown in green, and estimated recombination rates from HapMap are shown in light blue.

Cohort-specific association results for SNPs with the smallest *P* value from each locus implicated at or near genome-wide significance are shown in **Supplementary Table 3**.

For FEV$_1$, genome-wide significant associations were observed for 46 SNPs on chromosome 4q24 in or near four adjacent genes (**Supplementary Table 4**). The SNP with the smallest *P* value, rs17331332 ($P = 4.00 \times 10^{-10}$), is located near *NPNT*. The 45 other
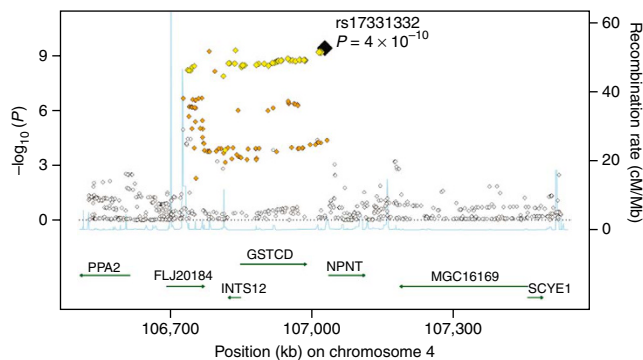

Figure 3 Regional association plot for the chromosome 4q24 locus associated with FEV$_1$ in the CHARGE consortium at genome-wide significance, which includes *FLJ20184*, *INTS12*, *GSTCD* and *NPNT*. Correlations between the target SNP (the SNP with the lowest *P* value, depicted in black) and other SNPs in the region are depicted in red when $r^2 = 1$, blue when $0.8 \leq r^2 < 1$, yellow when $0.5 \leq r^2 < 0.8$, orange when $0.2 \leq r^2 < 0.5$ and white when $r^2 < 0.2$. The r^2 values were based on the HapMap CEU population. Gene annotations are shown in green, and estimated recombination rates from HapMap are shown in light blue.

significantly associated SNPs include four SNPs located near the 5′ end of *NPNT*, 5 SNPs located in *INTS12* or near its 3′ end, seven SNPs located in *FLJ20184* or near its 3′ end and 29 SNPs located in *GSTCD*. *FLJ20184* encodes a hypothetical protein, according to several genome browsers including the UCSC Genome Browser (see URLs)[16], but there is no approved Human Gene Organization (HUGO) gene name for this locus[17]. The SNP rs17331332 is correlated at $r^2 > 0.5$ with most other significantly associated SNPs in this region (**Fig. 3**), suggesting that the associations in the four adjacent genes represent one independent finding. The significantly associated SNPs had MAFs between 6% and 8%. The absolute β (per-allele change) values for FEV$_1$ ranged from 55.92 to 71.43 ml (**Supplementary Table 4**), and the β directions were consistent across the CHARGE cohorts for all 46 genome-wide significant SNPs (**Supplementary Table 3** for rs17331332). Among these 46 SNPs, 39 were associated with FEV$_1$ at genome-wide significance among ever-smokers (**Supplementary Table 4**).

To evaluate whether other loci may also influence pulmonary function, we created Q-Q plots for FEV$_1$/FVC and FEV$_1$ among all participants after removing SNPs at or close to genome-wide significance and nearby SNPs correlated at $r^2 > 0.2$ with the top SNP for each locus (totaling 1,862 SNPs removed for FEV$_1$/FVC and 284 SNPs removed for FEV$_1$). The resulting Q-Q plots show some excess of small *P* values for FEV$_1$/FVC (**Supplementary Fig. 4a**) and FEV$_1$ (**Supplementary Fig. 4b**) over expectation.

Putative functional polymorphisms

Three SNPs among the 119 genome-wide significant SNPs for FEV$_1$/FVC are nonsynonymous (missense) polymorphisms: rs11155242

Table 2 Joint meta-analysis of SNPs selected from the top 8 loci implicated for FEV$_1$/FVC in the CHARGE GWAS and tested for replication with FEV$_1$/FVC in the SpiroMeta consortium[15]

SNP	Chr.	Gene	Ref. allele	CHARGE GWAS			SpiroMeta replication			Joint meta-analysis	
				Allele freq.[a]	β (%)	P	Allele freq.[a]	β (%)	P	β (%)	P
rs1980057	4	*HHIP*	T	0.40	0.51	**4.90 × 10⁻¹¹**	0.45	0.54	**1.09 × 10⁻¹⁰**	0.52	**3.21 × 10⁻²⁰**
rs1032295	4	*HHIP*	T	0.58	−0.47	**6.28 × 10⁻⁹**	0.55	−0.46	**1.15 × 10⁻⁷**	−0.47	**4.37 × 10⁻¹⁵**
rs3817928	6	*GPR126*	A	0.78	−0.59	**2.60 × 10⁻¹⁰**	0.79	−0.21	3.99 × 10⁻²	−0.42	**1.17 × 10⁻⁹**
rs7776375	6	*GPR126*	A	0.71	−0.51	**1.33 × 10⁻⁹**	0.73	−0.19	4.87 × 10⁻²	−0.37	**6.71 × 10⁻⁹**
rs6937121	6	*GPR126*	T	0.71	−0.49	**2.46 × 10⁻⁹**	0.72	−0.18	5.62 × 10⁻²	−0.35	**1.25 × 10⁻⁸**
rs11155242	6	*GPR126*	A	0.80	−0.54	**9.13 × 10⁻⁹**	0.80	−0.16	1.26 × 10⁻¹	−0.37	1.45 × 10⁻⁷
rs2277027	5	*ADAM19*	A	0.71	0.49	**8.32 × 10⁻¹⁰**	0.66	0.24	5.04 × 10⁻³	0.38	**9.93 × 10⁻¹¹**
rs1422795	5	*ADAM19*	T	0.66	0.48	**1.16 × 10⁻⁹**	0.66	0.24	6.65 × 10⁻³	0.37	**2.62 × 10⁻¹⁰**
rs2070600	6	*AGER*[b]	T	0.04	1.06	**1.49 × 10⁻⁸**	0.06	0.94	**4.40 × 10⁻⁷**	1.00	**3.15 × 10⁻¹⁴**
rs10947233	6	*PPT2*[b]	T	0.04	1.14	**4.71 × 10⁻⁸**	0.04	1.05	**3.43 × 10⁻⁵**	1.10	**6.66 × 10⁻¹²**
rs2869967	4	*FAM13A*	T	0.61	0.44	**1.57 × 10⁻⁸**	0.59	0.13	1.28 × 10⁻¹	0.30	1.91 × 10⁻⁷
rs6830970	4	*FAM13A*	A	0.65	0.46	**1.92 × 10⁻⁸**	0.64	0.11	2.00 × 10⁻¹	0.30	6.63 × 10⁻⁷
rs16909898	9	*PTCH1*	A	0.90	0.77	**1.78 × 10⁻⁸**	0.89	0.20	1.70 × 10⁻¹	0.50	5.34 × 10⁻⁷
rs10512249	9	*PTCH1*	A	0.10	−0.73	**2.79 × 10⁻⁸**	0.10	−0.19	1.67 × 10⁻¹	−0.48	5.75 × 10⁻⁷
rs1435867	2	*PID1*	T	0.93	−0.84	**3.74 × 10⁻⁸**	0.93	−0.075	6.51 × 10⁻¹	−0.50	1.53 × 10⁻⁵
rs10498230	2	*PID1*	T	0.07	0.86	**3.87 × 10⁻⁸**	0.07	0.075	6.56 × 10⁻¹	0.50	1.46 × 10⁻⁵
rs11168048	5	*HTR4*	T	0.58	−0.43	5.37 × 10⁻⁸	0.58	−0.36	**3.97 × 10⁻⁵**	−0.40	**1.08 × 10⁻¹¹**
rs7735184	5	*HTR4*	T	0.40	0.40	1.74 × 10⁻⁷	0.41	0.34	**5.62 × 10⁻⁵**	0.37	**6.23 × 10⁻¹¹**

'Gene' indicates gene either harboring the SNP or nearest to it. SNPs are grouped together by the nearest gene and ordered by the CHARGE GWAS P value. P values highlighted in bold exceeded the threshold for significance (P < 5 × 10⁻⁸ for the GWAS and joint meta-analysis, P < 8.33 × 10⁻⁴ for replication). β, per-allele change in FEV$_1$/FVC. Chr., chromosome. [a]Weighted average reference allele frequency for combined cohorts. [b]*AGER* and *PPT2* SNPs are considered to represent one locus given their correlations. FEV$_1$/FVC, forced expiratory volume in 1 s to forced vital capacity ratio.

(resulting in a lysine-to-glutamine substitution) in *GPR126*, rs1422795 (serine-to-glycine substitution) in *ADAM19* and rs2070600 (glycine-to-serine substitution) in *AGER*. The Polymorphism Phenotyping (PolyPhen) program (see URLs)[18] predicts that the amino acid substitutions resulting from rs11155242 and rs1422795 cause benign changes but that rs2070600 has a possibly damaging impact on the structure and function of AGER.

All other SNPs implicated for FEV$_1$/FVC or FEV$_1$ are intergenic, intronic or located in 3′ untranslated regions. Of these, three intronic *GPR126* SNPs (rs9496346, rs1040525 and rs6929442) and one intergenic SNP near *NPNT* (rs10516529) are located in transcription factor binding sites, according to the UCSC Genome Browser (see URLs)[16].

Replication with the SpiroMeta consortium

Thirty high-signal SNPs associated with FEV$_1$/FVC (18 SNPs from eight loci) or FEV$_1$ (12 SNPs from three loci) at or close to genome-wide significance were tested for replication in the SpiroMeta consortium, with results reported in a companion paper[15]. We evaluated these SNPs in 16,178 SpiroMeta participants of European ancestry with complete quantitative smoking data using the CHARGE analytic method (see statistical analysis section of Online Methods), which included adjustment for smoking status and pack-years, and performed joint meta-analyses of CHARGE GWAS and SpiroMeta replication results (**Tables 2** and **3**). P values that exceeded the significance threshold in SpiroMeta (P < 8.33 × 10⁻⁴ based on 60 tests)

Table 3 Joint meta-analysis of SNPs selected from the top three loci implicated for FEV$_1$ in the CHARGE GWAS and tested for replication with FEV$_1$ in the SpiroMeta consortium[15]

SNP	Chr.	Gene	Ref. allele	CHARGE GWAS			SpiroMeta replication			Joint meta-analysis	
				Allele freq.[a]	β	P	Allele freq.[a]	β	P	β	P
rs17331332	4	*NPNT*[b]	A	0.08	60.35	**4.00 × 10⁻¹⁰**	0.07	52.11	**2.45 × 10⁻⁶**	56.79	**5.69 × 10⁻¹⁵**
rs17036341	4	*NPNT*[b]	C	0.93	−58.85	**6.29 × 10⁻¹⁰**	0.94	−53.82	**6.38 × 10⁻⁷**	−56.65	**2.18 × 10⁻¹⁵**
rs11727189	4	*INTS12*[b]	T	0.06	63.43	**5.28 × 10⁻¹⁰**	0.05	66.38	**1.60 × 10⁻⁸**	64.70	**4.66 × 10⁻¹⁷**
rs17036090	4	*INTS12*[b]	T	0.93	−56.6	**1.37 × 10⁻⁸**	0.94	−59.51	**7.85 × 10⁻⁸**	−57.90	**5.61 × 10⁻¹⁵**
rs17036052	4	*FLJ20184*[b]	T	0.06	71.43	**6.07 × 10⁻¹⁰**	0.04	68.19	**5.78 × 10⁻⁷**	70.08	**1.83 × 10⁻¹⁵**
rs17035960	4	*FLJ20184*[b]	T	0.07	56.9	**3.87 × 10⁻⁹**	0.06	49.96	**4.52 × 10⁻⁶**	53.85	**9.42 × 10⁻¹⁴**
rs11097901	4	*GSTCD*[b]	T	0.07	58.85	**1.52 × 10⁻⁹**	0.06	59.49	**4.03 × 10⁻⁸**	59.14	**3.26 × 10⁻¹⁶**
rs11728716	4	*GSTCD*[b]	A	0.07	57.52	**1.80 × 10⁻⁹**	0.06	57.41	**7.61 × 10⁻⁸**	57.47	**7.20 × 10⁻¹⁶**
rs3749893	6	*TSPYL4*[c,d]	A	0.37	−26.55	5.35 × 10⁻⁷	0.37	4.01	4.62 × 10⁻¹	−11.71	2.05 × 10⁻³
rs1052443	6	*NT5DC1*[c,d]	A	0.62	26.12	7.50 × 10⁻⁷	0.62	−3.75	4.90 × 10⁻¹	11.61	2.17 × 10⁻³
rs6555465	5	*ADCY2*[d]	A	0.18	−31.23	1.32 × 10⁻⁶	0.19	0.12	9.86 × 10⁻¹	−16.24	4.96 × 10⁻⁴
rs7710510	5	*ADCY2*[d]	T	0.19	−30.34	1.72 × 10⁻⁶	0.21	−2.77	6.77 × 10⁻¹	−17.20	1.77 × 10⁻⁴

'Gene' indicates gene either harboring the SNP or nearest to it. SNPs are grouped together by the nearest gene and ordered by the CHARGE GWAS P value. P values highlighted in bold exceeded the threshold for significance (P < 5 × 10⁻⁸ for the GWAS and joint meta-analysis, P < 8.33 × 10⁻⁴ for replication). β, per-allele change in FEV$_1$ (mL). FEV$_1$, forced expiratory volume in one second. [a]Weighted average reference allele frequency for combined cohorts. [b]*NPNT*, *INTS12*, *FLJ20184*, and *GSTCD* SNPs are considered to represent one locus given their correlations. [c]*TSPYL4* and *NT5DC1* SNPs are considered to represent one locus given their correlations. [d]*TSPYL4*, *NT5DC1* and *ADCY2* SNPs were not associated with FEV$_1$ at genome-wide significance, but these loci had the next-smallest P values for association.

or the genome-wide significance threshold in joint meta-analyses ($P < 5 \times 10^{-8}$) were considered significant evidence for replication at their corresponding SNPs.

For FEV_1/FVC, among 18 SNPs tested for replication, six SNPs in three loci were significantly associated with this measure in SpiroMeta: rs1980057 and rs1032295 near *HHIP* ($r^2 = 0.72$), rs2070600 in *AGER* and rs10947233 in *PPT2* ($r^2 = 0.66$), and rs11168048 and rs7735184 in *HTR4* ($r^2 = 0.93$) (**Table 2**). Their joint meta-analysis *P* values ranged from 3.21×10^{-20} to 6.23×10^{-11} (**Table 2**). Five additional SNPs in *GPR126* (rs3817928, rs7776375 and rs6937121) and *ADAM19* (rs2277027 and rs1422795) were not significantly associated with FEV_1/FVC at genome-wide significance in SpiroMeta alone, but these SNPs were associated at genome-wide significance in the joint meta-analysis, with *P* values ranging from 9.93×10^{-11} to 1.25×10^{-8} (**Table 2**). For replicated SNPs, the allele frequencies and the direction and magnitude of the associations with FEV_1/FVC were similar between consortia (**Table 2**). Further, the *HHIP*, *ADAM19* and *HTR4* SNPs were significantly associated with FEV_1 in SpiroMeta (**Supplementary Table 5**). The *HHIP* SNP rs1980057 and the *HTR4* SNPs rs11168048 and rs7735184 were also associated with FEV_1 at genome-wide significance in the joint meta-analysis (*P* ranging from 5.86×10^{-9} to 1.58×10^{-8}, **Supplementary Table 5**). SNPs in *FAM13A*, *PTCH1* and *PID1* that gave genome-wide significance in CHARGE were not confirmed in analyses with SpiroMeta.

For FEV_1, among the 12 SNPs tested for replication, eight SNPs from one locus with four adjacent genes were significantly associated with this measure in SpiroMeta, including rs17331332 and rs17036341 near *NPNT*, rs11727189 and rs17036090 in or near *INTS12*, rs17036052 and rs17035960 in or near *FLJ20184*, and rs11097901 and rs11728716 in *GSTCD* (**Table 3**). For replicated SNPs, the allele frequencies and the direction and magnitude of the associations with FEV_1 were similar between consortia, and *P* values from joint meta-analysis ranged from 4.66×10^{-17} to 9.42×10^{-14} (**Table 2**). None of these SNPs were significantly associated with FEV_1/FVC in CHARGE or SpiroMeta (**Supplementary Table 5**).

Associations in individuals with normal pulmonary function

To address whether the genetic associations hold even among people with normal pulmonary function, we repeated the meta-analyses after excluding individuals with asthma or COPD, leaving 17,855 individuals ($n = 6,912$ from ARIC, $n = 2,634$ from CHS, $n = 6,371$ from FHS, $n = 1,126$ from RS-I and $n = 812$ from RS-II). Asthma was defined by self-report of ever having asthma or self-report of ever having physician-diagnosed asthma. COPD was defined spirometrically as having both FEV_1/FVC and FEV_1 less than the lower limit of normal values using National Health and Nutrition Examination Survey III prediction equations[19,20]. Comparing the original meta-analyses to the meta-analyses with exclusions for asthma and COPD, β estimates were highly correlated for the high-signal SNPs tested for replication (Pearson's $r > 0.99$ for 18 FEV_1/FVC SNPs and 12 FEV_1 SNPs). β estimates remained highly correlated for SNPs with *P* values as high as 0.01 in the original meta-analyses ($r = 0.92$ for FEV_1/FVC and $r = 0.96$ for FEV_1). As expected, there was some attenuation in *P* values for many of the SNPs in our implicated loci given the substantial power loss due to both reduced sample size and the truncation of the FEV_1/FVC and FEV_1 distributions, but there was substantial overlap in the top-ranking SNPs between the two meta-analyses (data not shown). The *P* values for some top-ranking SNPs became smaller, including several *ADAM19*, *FAM13A* and *HTR4* SNPs associated with FEV_1/FVC. Of note, 12 SNPs in *HTR4*, a locus with one SNP, rs11168048, showing borderline genome-wide significance in the original meta-analysis showed

genome-wide significance in the subset of individuals without asthma or COPD ($P = 6.93 \times 10^{-9}$ for rs11168048).

DISCUSSION

In meta-analyses of GWAS results in 20,890 CHARGE participants of European ancestry, we identified genome-wide significant associations with FEV_1/FVC for SNPs in seven previously unrecognized independent loci (*GPR126*, *ADAM19*, *AGER-PPT2*, *FAM13A*, *PTCH1*, *PID1* and *HTR4*) and with FEV_1 for one previously unrecognized independent locus annotated by at least three genes (*INTS12-GSTCD-NPNT*). The SpiroMeta consortium independently reported genome-wide significant associations of *GSTCD*, *HTR4*, *AGER*, *TNS1* and *THSD4* with FEV_1/FVC and FEV_1 (companion paper in this issue, ref. 15). Both consortia confirm previous GWAS findings implicating the *HHIP* region for FEV_1/FVC[10].

Several SNPs near the gene encoding the hedgehog interacting protein (*HHIP*) were associated with FEV_1/FVC at genome-wide significance in CHARGE and SpiroMeta, confirming earlier GWAS findings in FHS[10]. The hedgehog (Hh) signaling pathway is crucial in several embryonic development processes, including the branching morphogenesis of the lung[21,22]. Furthermore, several polymorphisms in three genes in the Hh signaling pathway (*IHH*, *HHIP* and *PTCH1*) were significantly associated with adult height in a previous GWAS[23]. Several *PTCH1* SNPs were also significantly associated with FEV_1/FVC in CHARGE, but these associations were not confirmed in SpiroMeta[15]. Epithelial cells produce Hh protein, which binds to its membrane receptor (encoded by *PTCH1*) on mesenchymal cells and orchestrates tissue and organ patterning. Hh pathway dysfunction during the fetal stage in humans is responsible for severe lung malformations[24,25]. In adults, the Hh-signaling pathway may participate in the response of the airway epithelium to injury, such as smoking and hyperoxia[26,27].

A nonsynonymous *AGER* SNP (rs2070600) was associated with FEV_1/FVC at genome-wide significance in our study and independently confirmed in SpiroMeta[15]. The AGER protein, a membrane-bound or soluble pattern-recognition receptor, belongs to the immunoglobulin superfamily of cell surface receptors. The SNP rs2070600 has functional significance: for example, it promotes higher ligand affinity and the production of proinflammatory proteins upon activation[28]. In healthy adult mice and humans, AGER is highly expressed in the lungs[29], and its absence contributes to the pathogenesis of idiopathic pulmonary fibrosis[30,31]. AGER signaling is involved in host defense, inflammation and tissue remodeling, processes that are relevant to accelerated decline in pulmonary function with age.

Polymorphisms in *HTR4* were associated with FEV_1/FVC at genome-wide significance in the joint meta-analysis of CHARGE and SpiroMeta results. *HTR4* encodes a G-coupled transmembrane receptor that regulates cyclic AMP production in response to 5-hydroxytryptamine (serotonin). Elevated levels of free serotonin have been found in the plasma of individuals with symptomatic asthma[32], and serotonin signaling pathways involving *HTR4* have been implicated in cholinergic and immune-mediated airway reactivity[33,34]. Upon activation by serotonin, *HTR4* in human airway epithelial cells regulates the release of a proinflammatory cytokine, a signature characteristic of asthma[35].

ADAM19 SNPs were associated with FEV_1/FVC at genome-wide significance in CHARGE and in the joint meta-analysis with SpiroMeta. *ADAM19* is a member of 'a disintegrin and metalloprotease' (ADAM) family of membrane-anchored glycoproteins that control cell-matrix interactions and help regulate growth and

morphogenesis. Polymorphisms in a gene that encodes for another ADAM family member, *ADAM33*, have been associated with bronchial hyper-responsiveness and accelerated lung function decline in those with asthma and in the general population[36–38]. *ADAM19* has not been previously implicated in human pulmonary disorders, but it is abundantly expressed in alveolar epithelial cells and bronchial smooth muscle tissue[39].

GPR126 polymorphisms were associated with FEV_1/FVC at genome-wide significance in CHARGE and in the joint meta-analysis with SpiroMeta. *GPR126* belongs to a superfamily of G protein–coupled receptors involved in cell adhesion and signaling[40]. Although the precise function of this gene has not been elucidated, its expression in mice is temporally increased during embryonic organ development and is highest in the adult lungs[41]. In humans, recent GWAS have linked *GPR126* variants with adult height, and more specifically with trunk height[42–44]. We adjusted all analyses for standing height. Therefore, we repeated analyses after adjusting for sitting height (a more reliable indicator of trunk height) for *GPR126* SNPs in ARIC, where both height variables were measured, and associations with FEV_1/FVC remained significant (data not shown). Thus, these associations are not likely to be due to residual confounding by trunk height.

Genome-wide significant associations with FEV_1 were observed in CHARGE for numerous SNPs spanning at least three genes on chromosome 4q24, and these associations were significant for all eight SNPs tested for replication in SpiroMeta (**Table 3**). There is moderate to strong linkage disequilibrium among the chromosome 4q24 SNPs, and the specific genes influencing FEV_1 remain speculative. The genes are ordered *INTS12-GSTCD-NPNT* along chromosome 4q24, and joint meta-analysis with SpiroMeta showed that SNPs from the genes *INTS12* and *GSTCD* had the most significant associations with FEV_1 (**Table 3**). The product of *INTS12* is a subunit of the integrator complex that associates with the C-terminal domain of RNA polymerase II and mediates 3′-end processing of small nuclear RNAs[45]. The glutathione S-transferase C-terminal domain (GSTCD) could influence lung function via mechanisms involving the detoxification by glutathione S-transferases of xenobiotics that might damage the lungs.

The most distal gene in the chromosome 4q24 region, *NPNT,* encodes nephronectin, which is expressed in fetal and adult lungs[46,47]. The *NPNT* SNP rs10516529 is located in a binding site for the transcription factor POU6F1 (also known as mPOU homeobox protein), which is known to be expressed in adult lungs and hypothesized to play a role in lung development[48–50]. A fourth predicted gene in the region, *FLJ20184*, is located proximal to the other three genes. Although *FLJ20184* encodes a hypothetical protein of unknown function, *FLJ20184* contains allelic variants associated with successful smoking cessation in a GWAS of patients in smoking cessation trials[51].

The genetic factors identified here gave estimated effect sizes consistent with those for well-established risk factors for pulmonary function decline. Carrying one copy of an implicated reference allele resulted in a FEV_1 difference ranging from 50–70 mL. These effect sizes correspond to approximately 2.8–3.9 years of age-related decline in pulmonary function based on a mean decline of about 18 mL/year and to approximately 1.7–2.3 years of active smoking-related decline based on a mean decline of about 30 mL/year[52]. Second-hand smoke exposure has also been associated with decline in FEV_1 (15 mL decline for a 10-year exposure in the home and 41 mL decline for a 10-year workplace exposure)[53]. For FEV_1/FVC, carrying one copy of an implicated reference allele resulted in a difference ranging from 0.30% to 1%. The lower effect-size estimates are comparable with the mean FEV_1/FVC decline related to second-hand smoking (0.35 for a 10-year exposure in the home and 0.14 for a 10-year workplace exposure)[53].

These comparisons demonstrate that the identified genetic factors have a moderate impact on pulmonary function. Individuals carrying these polymorphisms will have lower pulmonary function than predicted at a given age, thus placing them at greater risk for developing COPD and at a greater risk of mortality[2,3].

A GWAS of COPD identified *CHRNA3-CHRNA5* on chromosome 15 as a susceptibility locus[11]. *CHRNA3-CHRNA5* has also been associated with nicotine dependence[12,13]. In CHARGE, one identified SNP in this locus (rs1051730) was associated with FEV_1/FVC ($P = 0.00070$) and FEV_1 ($P = 0.016$), whereas the other identified SNP in this locus (rs8034191) was not associated with FEV_1/FVC ($P = 0.11$) or FEV_1 ($P = 0.36$). The nominal evidence for replication may reflect differences in study design and a potential gene-environment interaction involving smoking.

Our study has several important strengths. The CHARGE cohorts are well phenotyped with pulmonary function measures passing stringent quality control criteria, thus minimizing measurement error. Our large sample size of 20,890 participants offers a powerful resource to examine associations of common SNPs with modest to large effects[14]. Testing our most significant results in the SpiroMeta consortium provided independent replication of these associations. However, even with the large sample sizes in these combined consortia, we likely have insufficient power to detect associations of polymorphisms with small effect sizes or low frequencies.

Population-based cohorts are subject to population stratification, and analytic steps were taken to minimize this potential bias. Cohort-specific λ_{gc} values were low (1.00–1.05), and a genomic control adjustment was made in the meta-analyses to reduce inflation in the test statistics. The two largest cohorts, with the largest (albeit modest) λ_{gc} values (ARIC and FHS), incorporated principal components as potential confounders in their cohort-specific association tests. Although we cannot eliminate the possibility that some findings are subject to residual confounding by population stratification, the Q-Q plots showing deviations between observed and expected *P* values for many high- to moderate-signal SNPs and the replication of association for multiple top loci in SpiroMeta suggest a multifactorial influence on pulmonary function.

Our study identified several previously unrecognized loci related to two clinically important pulmonary function measures with evidence for replication, including *GPR126*, *ADAM19*, *AGER-PPT2* and *HTR4* for FEV_1/FVC and *INTS12-GSTCD-NPNT* for FEV_1 and confirmed previous reports of association with FEV_1/FVC in the *HHIP* region. These loci include genes with biologically plausible functions, and their identification here encourages future investigations to examine the mechanisms underlying their influence on pulmonary function. Fine mapping of these regions is needed to identify and characterize functional variants. Understanding the genetic determinants of pulmonary function is paramount in identifying the biological mechanisms that lead to its decline and in ultimately lessening the mortality burden associated with reduced pulmonary function.

METHODS

Methods and any associated references are available in the online version of the paper at http://www.nature.com/naturegenetics/.

Note: Supplementary information is available on the Nature Genetics website.

AUTHOR CONTRIBUTIONS

ARIC: D.B.H., L.R.L., N.F., M.B.S., D.J.C., N.M.P., A.C.M., K.E.N. and S.J.L. **CHS**: S.A.G., K.D.M., R.G.B., B.M.P., J.I.R., P.L.E., S.R.H. and T.L. **FHS**: J.B.W., T.-h.C. and G.T.O. **RS**: M.E., Y.M.T.A.v.D., G.G.B., C.M.v.D., A.G.U., A.H., F.R. and B.H.C.S. **Study design**: T.L., B.H.C.S., G.T.O. and S.J.L. **Data analysis**: D.B.H., M.E., J.B.W., L.R.L., K.D.M., N.F. and T.-h.C. **Drafting of manuscript**: D.B.H., M.E., J.B.W. and S.A.G. **Critical revision of manuscript**: D.B.H., M.E., J.B.W., S.A.G., L.R.L., K.D.M., N.F., Y.M.T.A.v.D., T.-h.C., R.G.B., M.B.S., D.J.C., G.G.B., B.M.P., C.M.v.D., J.I.R., A.G.U., A.H., N.M.P., F.R., A.C.M., P.L.E., K.E.N., S.R.H., T.L., B.H.C.S., G.T.O. and S.J.L.

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ONLINE METHODS

Pulmonary function measurements. Study design details of the participating CHARGE cohorts are described elsewhere[14,54–59]. Study protocols were approved by the relevant institutional review boards, and all participants provided written informed consent.

Pulmonary function testing was conducted by trained spirometry technicians at a single visit for RS and at more than one visit for ARIC, CHS and FHS. FEV_1/FVC and FEV_1 measures meeting American Thoracic Society or European Respiratory Society criteria for acceptability were tested for association with SNPs in participants of European ancestry who were successfully genotyped and provided informed consent for genetic testing.

In ARIC and CHS, pulmonary function measures and questionnaire data from the baseline visit were analyzed. ARIC measurements were made with a Collins Survey II water-seal spirometer (Collins Medical, Inc.) and Pulmo-Screen II software (PDS Healthcare Products, Inc.)[60]. CHS measurements were made with a Collins Survey I water-seal spirometer (Collins Medical, Inc.) and software from S&M Instruments[61,62].

In three generations of families participating in the FHS, data from the most recent examination were analyzed. Eligible examinations providing spirometry and questionnaire data included examinations 13, 16, 17 and 19 in the original cohort (in approximate 2-year intervals); examinations 3, 5, 6 and 7 in the offspring generation (in approximate 4-year intervals); and the one examination completed to date for the third generation. Equipment used in the standard protocol evolved as technology improved over the decades of study[63]. A Collins Survey water-filled spirometer (Collins Medical, Inc.) was used for most examinations, with measurements made by Eagle II microprocessor (Collins Medical, Inc.) or by software from the S&M Instruments. In more recent examinations, a Collins Comprehensive Pulmonary Laboratory dry rolling-seal spirometer and Collins 2000 Plus/SQL Software (Collins Medical, Inc.) were used.

In RS, pulmonary function was measured at the fourth center visit of participants from the original cohort (RS-I) and the second center visit of participants from the first extension cohort (RS-II). Spirometry was performed using a SpiroPro portable spirometer (Erich Jaeger GmbH)[64,65].

Genotyping, imputation and quality control. Different genotyping platforms were used across the cohorts[14] (**Table 1**). Imputation was conducted using either MACH[66] or BIMBAM[67] to generate approximately 2.5 million autosomal SNP genotype dosages for meta-analysis. The imputation methods perform similarly, although MACH generally produces higher accuracy rates than the imputation process used in BIMBAM (fastPHASE)[68]. Differing imputation methods across cohorts is not a source of bias for meta-analysis because all comparisons using the imputed data are within-cohort comparisons.

ARIC. Among 8,861 self-identified white ARIC participants genotyped, 8,127 participants remained in the study after exclusions for call rate <95%, genotypic and phenotypic sex mismatch, discordances with previous genotype data, suspected first-degree relative of an included individual based on genotype data, more than 8 s.d. for any of the first ten principal components using EIGENSTRAT[69], or outlying average identity-by-state estimates using PLINK[70]. Of these, 7,980 participants had available pulmonary function measures and complete covariate information.

A total of 704,588 autosomal genotyped SNPs remained after exclusions for call rate < 95%, MAF < 1%, Hardy-Weinberg equilibrium (HWE) $P < 10^{-5}$ or lack of strand annotation. MACH (version 1.00.16)[66] was used to impute all autosomal SNPs with reference to HapMap CEU (release 21, build 35)[71] from these 704,588 SNPs. Imputed SNPs failing additional quality control criteria (monomorphism, HWE $P < 10^{-6}$, or genotype frequencies between two genotyping phases differing by $P < 10^{-6}$) were excluded, leaving 2,515,866 genotyped or imputed SNPs for analysis.

CHS. CHS genotyped 3,980 participants free of cardiovascular disease at baseline with available DNA and consent to genetic testing. After exclusions for call rate < 95%, sex mismatch or discordance with prior genotyping, 3,291 self-identified white participants remained. Of these, 3,140 had pulmonary function measures and complete covariate information.

A set of 306,655 autosomal genotyped SNPs remained after exclusions for call rate <97%, HWE $P < 10^{-5}$, more than two duplicate errors or mendelian inconsistency (for reference HapMap CEU trios)[71], heterozygote frequency > 0

or no mapping in dbSNP. Imputation of autosomal SNPs was based on these 306,655 SNPs using BIMBAM (version 0.99)[67] with reference to HapMap CEU (release 22, build 36)[71]. The analysis dataset included 2,543,887 genotyped or imputed SNPs.

FHS. A total of 8,481 participants remained after exclusions for call rate < 97%, heterozygosity >5 s.d. from the mean, or excessive non-inheritance. The analysis dataset included 7,694 participants with complete spirometry and covariate data.

MACH (version 1.00.15)[66] was used for imputation based on 378,163 autosomal SNPs remaining after exclusions for HWE $P < 10^{-6}$, call rate < 97%, differential missingness related to genotype (mishap procedure in PLINK[70]) with $P < 10^{-9}$, mendelian errors > 100, MAF < 1% or absence from HapMap. Two hundred unrelated individuals with high call rates were used to infer model parameters, which were subsequently applied to all 8,481 individuals. Imputation, using HapMap CEU (release 22, build 36)[71], produced genotype dosages on 2,543,887 genotyped or imputed SNPs.

RS. All RS participants with available DNA were genotyped; 5,974 RS-I participants and 2,157 RS-II participants remained after exclusion for call rate < 97.5%, excess autosomal heterozygosity, sex mismatch or outlying identity-by-state clustering estimates. Of these, 1,224 RS-I participants and 852 RS-II participants had pulmonary function measures and complete covariate information.

After exclusions for call rate < 98%, HWE $P < 10^{-6}$, and MAF < 1%, 512,349 autosomal SNPs in RS-I and 466,389 autosomal SNPs in RS-II were used for imputation in MACH (version 1.00.15 for RS-I and 1.00.16 for RS-II)[66] with reference to the 2,543,887 SNPs of the HapMap CEU (release 22, build 36)[71].

Statistical analysis. In cross-sectional analyses, FEV_1/FVC and FEV_1 were tested for association with SNP genotypes using a 1-degree-of-freedom additive model of the dosage value (estimated reference allele count with a fractional value ranging from 0 to 2.0) as a predictor in linear regression models. Associations were examined overall and stratified into ever- and never-smokers. Overall models were adjusted for age, sex, standing height, smoking status (current, past or never-smoker) and pack-years of smoking. Current, past or never smoking was based on questionnaire responses, and pack-years were calculated for current and past smokers by multiplying smoking dose (packs per day) and duration (years smoked). Stratified models used the same covariates as the overall models, except that the ever-smoker stratum included adjustment for smoking status as current or past and the never-smoker stratum included no smoking-related covariates. Additional study-specific covariates included recruitment cohort (FHS), recruitment center (ARIC and CHS) and principal component eigenvalues for population stratification adjustments (ten components for ARIC and statistically significant components for FHS). Models were implemented using ProbABEL[72] in ARIC, R[73] in CHS, linear mixed effects models with fixed effects for SNPs and random effects for individuals correlated within families[74] in FHS, and MACH2QTL[66] in RS as implemented in GRIMP[75]. In FHS, the kinship package in R generated a covariance matrix for each family based on the kinship coefficient for each relative pair. The kinship matrix, which includes the full set of family-specific covariance matrices, specified the covariance matrix for the random effects.

GWAS results from the four cohorts were combined using inverse variance weighted meta-analysis in METAL (see URLs). Meta-analysis was performed on approximately 2,534,500 SNPs after applying genomic control for each study and filtering out SNPs with extremely low imputation quality ratios (<0.01) and MAFs (<1%).

The genome-wide significance threshold was defined a priori as $P < 5 \times 10^{-8}$, the Bonferroni adjustment for 1 million independent tests[76]. Information on SNP function and position relative to genes, microRNA and transcription factor binding sites was obtained using a Perl script (J.B.W.) that queries tables of the UCSC genome browser[16] (hg18, March 2006 genome build, see URLs). Functional effects of nonsynonymous SNPs on protein structure and function were predicted using PolyPhen (see URLs)[18].

Replication in the SpiroMeta consortium. We exchanged 30 SNPs for replication testing with the SpiroMeta consortium (companion manuscript[15]). No additional genotyping was required, as these SNPs were available from

the SpiroMeta GWAS. We aimed to select two SNPs from each of the top genes implicated for FEV_1/FVC or FEV_1, with nearly all the genes exceeding genome-wide significance. The SNP with the lowest P value in or near each gene was selected. A second SNP, genotyped (instead of imputed) in at least one cohort, was selected with preference for nonsynonymous SNPs and SNPs not in strong linkage disequilibrium with the first selected SNP. Only one SNP was available for *AGER*, *PPT2*, *TSPYL4* and *NT5DC1*. Four SNPs were selected from two linkage disequilibrium blocks for the largest gene, *GPR126*. In total, 18 SNPs from 9 genes (8 independent loci) implicated for FEV_1/FVC and 12 SNPs from 7 genes (3 independent loci) implicated for FEV_1 were tested for replication.

Unlike CHARGE, SpiroMeta used normalized residuals as phenotypes, adjusted for age^2 rather than age, and did not adjust for smoking. For better comparison, SpiroMeta conducted modified analyses following the CHARGE analytic method described above in 16,178 participants from adult cohorts with complete quantitative smoking data available. Results from the CHARGE GWAS and SpiroMeta replication were combined in a joint meta-analysis using inverse variance weighting with METAL. SpiroMeta results with $P < 8.33 \times 10^{-4}$, based on an overly conservative Bonferroni correction for 60 tests (30 SNPs tested for association with two traits, FEV_1/FVC and FEV_1), or joint meta-analysis results with $P < 5 \times 10^{-8}$ (genome-wide significance threshold), were considered statistically significant.

URLs. METAL, http://www.sph.umich.edu/csg/abecasis/metal/; UCSC Genome Browser, http://genome.ucsc.edu/; Polymorphism Phenotyping (PolyPhen), http://genetics.bwh.harvard.edu/pph/.

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